

# R-scape User's Guide

---

RNA Significant Covariation Above Phylogenetic Expectation

Version v2.5.7; March 2025

Elena Rivas  
elenarivas@fas.harvard.edu  
Department of Molecular and Cellular Biology  
Harvard University  
52 Oxford Street, Northwest Building #430  
Cambridge MA 02138 USA  
<http://rivaslab.org/>

Copyright (C) 2017-2025 Elena Rivas, Harvard University.

Permission is granted to make and distribute verbatim copies of this manual provided the copyright notice and this permission notice are retained on all copies.

R-scape is licensed and freely distributed under the GNU General Public License version 3 (GPLv3). For a copy of the License, see <http://www.gnu.org/licenses/>.

# Contents

<b>1</b>	<b>Introduction</b>	<b>5</b>
	How to avoid reading this manual . . . . .	5
	How do I cite R-scape? . . . . .	5
	How do I cite CaCoFold? . . . . .	5
	How do I cite CaCoFold-R3D? . . . . .	5
<b>2</b>	<b>Installation</b>	<b>6</b>
	Quick installation instructions . . . . .	6
	System requirements . . . . .	6
	Makefile targets . . . . .	7
	Why is the output of 'make' so clean? . . . . .	7
	What gets installed by 'make install', and where? . . . . .	7
<b>3</b>	<b>Tutorial for running R-scape</b>	<b>8</b>
	Modes of R-scape . . . . .	8
	The four options to run R-scape . . . . .	8
	Option <code>-RAF(S)</code> disallowed . . . . .	9
	Files used in the tutorial . . . . .	9
	Running R-scape on one alignment file . . . . .	10
	Default parameters . . . . .	11
<b>4</b>	<b>Tutorial for running CaCoFold</b>	<b>13</b>
	The <code>-cacofold</code> option . . . . .	13
	Example of an RNA with pseudoknots . . . . .	15
	Single sequence structure prediction . . . . .	18
<b>5</b>	<b>Tutorial for running CaCoFold-R3D</b>	<b>19</b>
	The <code>-cacofold -r3d</code> options . . . . .	19
	Examples of 3D motifs and how to incorporate them to a R3D descriptor file . . . . .	20
	Hairpin Loops (HL) . . . . .	20
	Bulge Loops (BL) . . . . .	21
	Internal Loops (IL) . . . . .	22
	3-way Junctions (J3) . . . . .	24
	4-way Junctions (J4) . . . . .	25
	Branch Segments (BS) . . . . .	26
	The <code>-r3dfile</code> to customize the descriptor file . . . . .	27
<b>6</b>	<b>Inputs</b>	<b>28</b>
	The Stockholm format to describe a consensus structure . . . . .	28
	A extended Stockholm format to describe consensus structures with overlapping base pairs . . . . .	28
<b>7</b>	<b>Outputs</b>	<b>29</b>
	Covariation tabular output . . . . .	29
	Power tabular output . . . . .	30
	Default graphical outputs . . . . .	30
	Details about graphical outputs . . . . .	31
	Using option <code>-cacofold</code> . . . . .	32
	Graphical outputs per alignment . . . . .	33

<b>8 Options</b>	<b>36</b>
Covariation statistic options	36
-E <x>	36
--GT, --MI, --MIr, --MIg, --CHI, --OMES, --RAF, --RAFS,	36
--C2, --C16, --CWC	37
Options to calculate power	37
--singlesubs	37
--joinsubs	37
--doublesubs	37
Covariation aggregation options	37
--fisher,--sidak	37
--lancaster,--wfisther	37
--lancaster_join,--wfisther_join	37
--lancaster_double,--wfisther_double	37
Search options	38
-s	38
--cacofold	38
--naive	38
--tstart <n>	38
--tend <n>	38
--window <n>	38
--slide <n>	38
--vshuffle	38
--cshuffle	38
--giventnull <f>	38
Input alignment options	39
-I <x>	39
--gapthresh <x>	39
--consensus	39
--submsa <n>	39
--treefile <f>	39
--ntree <n>	39
Options for producing a CaCoFold structure	39
--cacofold	39
--cacofold --r3d	39
--r3dfile <f>	39
--cyk	40
--decoding	40
--refseq	40
--E_neg <x>	40
--lastfold	40
--show.hoverlap	40
--covmin <n>	40
--allow.negatives	40
--Rfam	40
--rmcoding	40
Options for importing a structure	41
--pdb <s>	41
--cntmaxD <x>	42
--cntmind <n>	42
--onlypdb	42

--draw_nonWC . . . . .	42
Options for type of pairs tested . . . . .	43
--samplecontacts . . . . .	43
--sampleebp . . . . .	43
--samplewc . . . . .	43
Output options . . . . .	43
--roc . . . . .	43
--outmsa <f> . . . . .	44
--outtree <f> . . . . .	44
--savenull . . . . .	44
Plotting options . . . . .	44
--nofigures . . . . .	44
--r2rall . . . . .	44
Other options . . . . .	44
--seed <n> . . . . .	44
<b>9 Some other topics . . . . .</b>	<b>45</b>
How do I cite R-scape? . . . . .	45
How do I cite CaCoFold? . . . . .	45
How do I cite CaCoFold-R3D? . . . . .	45
How do I report a bug? . . . . .	45
<b>10 Acknowledgments . . . . .</b>	<b>47</b>

# 1 Introduction

R-scape (RNA Significant Covariation Above Phylogenetic Expectation) is a program that given a multiple sequence alignment (MSA) of RNA sequences, finds the pairs of positions that show a pattern of significant covariation. Each covariation score has an E-value associated to it. E-values are determined using a null model of covariation due to phylogeny but independent of any structural constraints.

## How to avoid reading this manual

- Follow the quick installation instructions on page 6.
- Go to the tutorial section on page 8, which walks you through some examples of using R-scape on real data.

Everything else, you can read later.

## How do I cite R-scape?

Rivas, E. *et al.*, “A statistical test for conserved RNA structure shows lack of evidence for structure in lncRNAs”, *Nature Methods* 14, 4548 (2017).

<https://www.nature.com/articles/nmeth.4066>

Rivas, E. and Eddy, S. E., “Response to Tavares *et al.*, Covariation analysis with improved parameters reveals conservation in lncRNA structures”, (2018).

<https://doi.org/10.1101/2020.02.18.955047>.

Rivas, E. *et al.*, “Estimating the power of sequence covariation analysis for detecting conserved RNA structure”, *Bioinformatics*, 36, 30723076, (2020).

<https://doi.org/10.1093/bioinformatics/btaa080>

Rivas, E., “RNA covariation at helix-level resolution for the identification of evolutionarily conserved RNA structure”, *PLOS Comput Biol*, 19(7): e1011262, (2023).

<https://doi.org/10.1371/journal.pcbi.1011262>.

## How do I cite CaCoFold?

Rivas, E., “RNA structure prediction using positive and negative evolutionary information”, *PLOS Comput Biol*, 16(10), e1008387, (2020).

<https://doi.org/10.1371/journal.pcbi.1008387>.

## How do I cite CaCoFold-R3D?

Karan, A and Rivas, E., “All-at-once RNA folding with 3D motif prediction framed by evolutionary information”, *bioRxiv*, Dec (2024).

<https://doi.org/10.1101/2024.12.17.628809>, 20 Dec 2024.

## 2 Installation

### Quick installation instructions

Download `R-scape.tar.gz` from <http://eddylab.org/>; unpack it, configure, and make:

```
> tar xf R-scape.tar.gz
> cd R-scape
> ./configure
> make
> make install
```

The newly compiled binary (`R-scape`) is in the `R-scape/bin` directory. You can run it from there, as in this example:

```
> bin/R-scape tutorial/updatedArisong.sto
```

That's it. You can keep reading if you want to know more about customizing a `R-scape` installation, or you can skip ahead to the next chapter, the tutorial.

### System requirements

**Operating system:** `R-scape` is designed to run on POSIX-compatible platforms, including UNIX, Linux and MacOS/X. The POSIX standard essentially includes all operating systems except Microsoft Windows. We have tested most extensively on Linux and MacOS/X because these are the machines we develop on.

**Compiler:** The source code is C conforming to POSIX and ANSI C99 standards. It should compile with any ANSI C99 compliant compiler, including the GNU C compiler `gcc`, and the C++ compiler `g++`. We test the code using the `gcc` and `g++` compilers.

The code include several Perl scripts (from the independent program `R2R` used here). Make sure your `PATH` environmental variable includes a directory with a Perl executable.

The code also uses `GNUPLOT`. Make sure your `PATH` environmental variable includes a directory with a `GNU-PLOT` executable.

**Libraries and other installation requirements:** `R-scape` includes three software libraries:

- the Easel library package (<http://bioeasel.org/>),
- the HMMER library package (<http://hmmer.org/>),
- the Infernal library package (<http://eddylab.org/infernal/>),

and three independent programs:

- FastTree (Price et al., 2010) (for building phylogenetic trees, v2.1.11),
- R2R (Weinberg and Breaker, 2011) (for drawing consensus RNA structures),
- RNAVIEW (Yang et al., 2003) (for identifying different types of basepairs in nucleic acid alignments).

All libraries and independent programs will automatically compile during `R-scape`'s installation process. By default, `R-scape` does not require any additional libraries to be installed by you, other than standard ANSI C99 libraries that should already be present on a system that can compile C code.

Executables for the three independent programs will appear in the `R-scape/bin` directory.

## Makefile targets

**all** Builds everything. Same as just saying `make`.

**install** Installs the binaries (`R-scape`, `FastTree`, `r2r`).

By default, programs are installed in `R-scape_version/bin`. You can customize the location of the binaries by replacing

```
> ./configure
```

with

```
> ./configure --prefix=/the/directory/you/want
```

The newly compiled binaries are now in the `/the/directory/you/want/bin` directory.

**uninstall** Reverses the steps of `make install`.

**clean** Removes all files generated by compilation (by `make`). Configuration (files generated by `./configure`) is preserved.

**distclean** Removes all files generated by configuration (by `./configure`) and by compilation (by `make`).

## Why is the output of 'make' so clean?

Because we're hiding what's really going on with the compilation with a wrapper. If you want to see what the command lines really look like, pass a `V=1` option (V for "verbose") to `make`, as in:

```
> make V=1
```

## What gets installed by 'make install', and where?

The top-level configure file has a variable `RSCAPE_HOME` that specifies the directory where `make install` will install things: `RSCAPE_HOME/bin`.

By default `RSCAPE_HOME` is assigned to the current directory `R-scape`.

The best way to change this default is when you use `./configure`, and the most important variable to consider changing is `--prefix`. For example, if you want to install `R-scape` in a directory hierarchy all of its own, you might want to do something like:

```
> ./configure --prefix=/usr/local/rscape
```

That would keep `R-scape` out of your system-wide directories like `/usr/local/bin`, which might be desirable. Of course, if you do it that way, you'd also want to add `/usr/local/rscape/bin` to your `$PATH`.

### 3 Tutorial for running R-scape

Here's a tutorial walk-through of how to use R-scape. This should suffice to get you started.

#### Modes of R-scape

For an input alignment, R-scape reports all pairs that have covariation scores with E-values smaller than a target E-value.

R-scape has two different **modes** of operation which determine how it calculates E-values, for which it needs to know how many possible base pairs were tested (i.e. E-values are multiple-test-corrected). The E-values are calculated in one of two ways:

**A one-set statistical test:** *default*

E-values are calculated assuming that all pairs are possible.

This is the default behavior of R-scape.

**A two-set statistical test:** `option -s`

If the alignment has associated a *given structure*, **option -s** performs two independent statistical tests: one for the pairs included in the structure, a different one for all the remaining possible pairs.

It also draws the given consensus structure annotated with the significantly covarying base pairs.

#### The four options to run R-scape

These are the four options to run R-scape.

Evaluate region for conserved structure	<p>All possible pairs are analyzed equally as a one set test. If a consensus structure is provided, that structure is ignored in the covariation test, but it is visualized with the significant covarying pairs highlighted in green.</p> <p><b>preferred use:</b> This option is most appropriate if you're trying to determine if a conserved structure exists.</p>
Predict new structure	<p>All possible pairs are analyzed equally. A structure is predicted and visualized with the significant covarying pairs highlighted in green.</p> <p><b>preferred use:</b> This option is most appropriate for obtaining a new consensus structure prediction based on covariation analysis.</p>
Evaluate given structure	<p>Requires that your Stockholm file has a proposed consensus structure annotation. Two independent covariation tests are performed, one on the set of proposed basepairs, the other on all other possible pairs. The given structure is visualized with significant covarying pairs highlighted in green.</p> <p><b>preferred use:</b> This option is most appropriate for evaluating how well an independently proposed consensus structure is supported by covariation analysis.</p>
Improve given structure	<p>Requires that your Stockholm file has a proposed consensus structure annotation. Two independent covariation tests are performed, one on the set</p>

of proposed basepairs, the other on all other possible pairs. A new consensus structure is predicted and visualized with the significant covarying pairs highlighted in green.

**preferred use:**

This option is most appropriate for using covariation analysis to improve your current consensus structure.

I'll show examples of running each mode, using examples in the `tutorial/` subdirectory of the distribution.

## Option `-RAF(S)` disallowed

The options to use the covariation measures `RAF`, `RAFa`, `RAFp`, `RAFS`, `RAFSp`, and `RAFSa` has been disallowed, unless they are used in combination with option `-naive` which reports the list of values for all possible pairs without any statistical significance associated to them.

The following disclaimer appears otherwise.

```
> bin/R-scape --RAF tutorial/updated_Arisong.sto
```

```
DISCLAIMER: This measure can only be used in combination with the --naive option.
```

```
The --naive option reports a ranked list of scores for all possible
pairs without assigning E-values. RAF, RAFS and related measures
(RAFp, RAFa, RAFSp, RAFSa) cannot be used in combination with
R-scape's statistical test.
```

```
The RAF(S) statistics measure covariation and consistency. RAFS
assigns relatively high scores to pairs of alignment columns that are
consistent with base pairing even if there is no covariation at
all. The RAFS statistic was developed for the purpose of predicting
consensus RNA structures from alignments of sequences already presumed
to have a structure (Hofacker et al., 2002; Lindgreen et al.,
2006). For this purpose, both covariation and consistency are useful
cues. Distinguishing a conserved RNA structure from a conserved
primary sequence is a different problem that requires using a
statistic that does not systematically detect significant signals on
conserved primary sequence alone. That is R-scape's statistical
test. The R-scape statistical test can only be used with measures that
estimate covariation alone such as mutual information (MI) or G-test
(GT).
```

## Files used in the tutorial

The subdirectory `/tutorial` in the R-scape distribution contains the files used in the tutorial.

The tutorial provides several examples of RNA structural alignments, all in Stockholm format:

**updated\_Arisong.sto** Structural alignment of the ciliate Arisong RNA. This alignment is an updated version of the one published in (Jung et al., 2011).

**ar14.sto** Structural alignment of the  $\alpha$ -proteobacteria ncRNA ar14. This alignment is an updated version of the one published in (del Val et al., 2012).

**manA.sto** Alignment of the manA RNA motif (?Weinberg et al., 2010) provided in the Zasha Weinberg database (ZWD) (Weinberg, 2018).

**RF00005.sto** Rfam v12.0 (Nawrocki et al., 2015) seed alignment of tRNA.

**RF00001-noss.sto** Rfam v12.0 seed alignment of 5S rRNA, after removing the consensus secondary structure.

## Running R-scape on one alignment file

To run R-scape with default parameters on alignment file `tutorial/updated_Arisong.sto` use:

```
> bin/R-scape tutorial/updated_Arisong.sto
```

The output is a list of the significantly covarying positions under the one-set test

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.4.0 (Oct 2019)
# Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
#-----
# MSA updated_Arisong_1 nseq 95 (95) alen 66 (150) avgid 65.82 (64.97) nbpairs 20 (20)
# One-set statistical test (all pairs are tested as equivalent)
#
#
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTp 0.05 [-9.78,121.66] [0 | 2 20 2 | 10.00 100.00 18.18]
#
# left_pos right_pos score E-value substitutions power
#-----
* 98 106 121.65645 0.00241628 45 0.48
* 122 137 91.44593 0.038356 57 0.58
```

A star “\*” in the first column indicates that the pair is part of the annotated structure in the `updated_Arisong.sto` file. A blank indicates a pair that is not compatible with the structure. A “~” indicates an interaction not in the annotated structure but compatible with it (none in this example).

The Arisong RNA in `tutorial/updated_Arisong.sto` has a proposed secondary structure. Instead of testing all pairs as equivalent, we may want to test the significance of the given structure as a one set of pairs, and independently that of the rest of all possible pairs. In order to do a two-set test use:

```
> bin/R-scape -s tutorial/updated_Arisong.sto
```

The output is a list of the significantly covarying positions under the two-set test.

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.4.0 (Oct 2019)
# Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
#-----
# MSA updated_Arisong_1 nseq 95 (95) alen 66 (150) avgid 65.82 (64.97) nbpairs 20 (20)
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
#
#
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTp 0.05 [-9.78,121.66] [0 | 11 20 11 | 55.00 100.00 70.97]
#
# left_pos right_pos score E-value substitutions power
#-----
* 98 106 121.65645 2.25295e-05 45 0.48
* 122 137 91.44593 0.000357632 57 0.58
* 96 108 88.43400 0.000466924 26 0.28
* 120 139 74.80289 0.00162024 87 0.76
* 119 140 58.72158 0.00678565 90 0.78
* 121 138 58.34837 0.00691674 99 0.82
* 94 110 57.27959 0.00760538 37 0.40
* 124 134 55.67692 0.0086606 20 0.21
* 123 135 54.59630 0.00946822 72 0.68
* 99 105 53.44797 0.0107226 15 0.14
* 97 107 44.91842 0.0405594 58 0.59

# The given structure
# SS_cons ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons ::::::::::::::::::::::::::::::::::::::::::::::<<<<<<_>>>>>><<<<<<--<<
#
# SS_cons <<<<<_>>>>>>-->>>>>>:::
#
```

```

# Power analysis of given structure
#
# covary   left_pos   right_pos   substitutions   power
#-----
#      *    94         110         37         0.40
#          95         109         28         0.31
#      *    96         108         26         0.28
#          97         107         58         0.59
#      *    98         106         45         0.48
#      *    99         105         15         0.14
#          100        104         20         0.21
#          111        148         0         0.00
#          112        147         18         0.18
#          113        146         1         0.00
#          114        145         15         0.14
#          115        144         49         0.52
#          116        143        106         0.84
#      *    119        140         90         0.78
#      *    120        139         87         0.76
#      *    121        138         99         0.82
#      *    122        137         57         0.58
#      *    123        135         72         0.68
#      *    124        134         20         0.21
#          125        133         31         0.34
#
# BPAIRS 20
# avg substitutions per BP  43.7
# BPAIRS expected to covary 8.3
# BPAIRS observed to covary 11

```

The scores of the pairs are identical to those in the one-set test. The E-values have changed relative to those of the one-set test.

## Default parameters

Default parameters are:

**Target E-value:** default is 0.05. R-scape reports pairs which covariation score has E-value smaller or equal to the target value. The target E-value can be changed with option **-E <x>**,  $x \geq 0$ .

**Sequence weighting:** Sequences are weighted according to the Gerstein/Sonnhammer/Chothia (GSC) algorithm (Gerstein et al., 1994). This algorithm is time consuming. For alignments with more than 1000 sequences, we use the faster position-based weighting algorithm (Henikoff and Henikoff, 1994). Both weighting algorithms are implemented as part of the easel library.

**Gaps in columns:** Columns with more than 50% gaps are removed. The gap threshold for removing columns can be modified using option **--gapthresh <x>**,  $0 < x \leq 1$ .

**Covariation statistic:** The default covariation statistic is the average product corrected G-Test (equivalent to option **--Gtp**).

**Covariation Class:** R-scape uses the 16 component covariation statistic (C16), unless the number of sequences in the alignment is  $\leq 8$  or the length of the alignment is  $\leq 50$ , in which case it uses the two-class covariation statistic (C2). A particular covariation class can be selected using either **--C16** or **--C2**.

The threshold for the minimum number of sequences can be changed with option **--nseqthresh <n>**. The threshold for the minimum alignment length can be changed with option **--alenthresh <n>**.

**Null alignments:** In order to estimate E-values, R-scape produces 20 null alignments, unless the product of the number of sequences by the length of the alignment  $< 10,000$  in which case the number of null alignments is 50; or  $< 1,000$  in which case it is 100. The number of null alignments can be controlled with option **--nshuffle** **<n>**.

A full list of the R-scape options is found by using

```
> R-scape -h
```

## 4 Tutorial for running CaCoFold

Here's a tutorial walk-through of how to use CaCoFold. CaCoFold uses the evolutionary information capture by R-scape in the form of the base pairs that significantly covary, and builds a structure. The CaCoFold structure may contain pseudoknots and triplets, provided that they present covariation signal.

### The `--cacofold` option

After performing one of the two statistical tests with R-scape, this option implements the CaCoFold algorithm:

Builds the best consensus structure that includes the largest possible number of significantly covarying pairs, *the maximum-covariation optimal consensus structure*. The algorithm identifies pseudoknots and other not nested interactions by running a cascade of nested algorithms until all covarying pairs are taken into account.

Draws the *maximum-covariation optimal consensus structure* annotated with the significantly covarying base pairs.

It also returns the alignment in Stockholm format annotated with the max-cov optimal consensus structure.

```
> bin/R-scape --cacofold tutorial/updated_Arisong.sto
```

The output includes first the same output as default R-scape alone, followed by R-scape's proposed structure that under the heading "# The predicted CaCoFold structure" as follows,

```
# The predicted CaCoFold structure
# SS_cons ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons ::::::::::::::::::::::::::::::::::::::::::<<<<_____>>>>:<<<<---<<
#
# SS_cons <<<<_____>>->>>>---->>>>::
#

# Power analysis of CaCoFold structure
#
# covary  left_pos      right_pos    substitutions      power
#-----
#          95           109           28              0.31
#          96           108           26              0.28
#          97           107           58              0.59
#      *    98           106           45              0.48
#          99           105           15              0.14
#          111          148            0              0.00
#          112          147           18              0.18
#          113          146            1              0.00
#          114          145           15              0.14
#          115          144           49              0.52
#          119          140           90              0.78
#          120          139           87              0.76
#          121          138           99              0.82
#      *    122          137           57              0.58
#          123          135           72              0.68
#          124          134           20              0.21
#
# BPAIRS 16
# avg substitutions per BP  42.5
# BPAIRS expected to covary 6.5
# BPAIRS observed to covary 2
```

The structure predicted by R-scape includes all the basepairs reported as covarying, provided that those can be arranged into one single structure (including pseudoknots and other non Watson-Crick interactions). The R-scape folding algorithm cannot deal with residues that covary with more than one other residue, such as is the case for alternative structures or triplets.

Similarly using

```
> bin/R-scape -s --cacofold tutorial/updated_Arisong.sto
```

The output includes first the same output as **option -s** of R-scape alone, followed by R-scape's proposed CaCoFild structure including all the the covarying pairs obtained under the two-set test.

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.4.0 (Oct 2019)
# Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
#-----
# MSA updated_Arisong_1 nseq 95 (95) alen 66 (150) avgid 65.82 (64.97) nbpairs 20 (20)
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
#
#
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTp 0.05 [-9.78,121.66] [0 | 11 20 11 | 55.00 100.00 70.97]
#
# left_pos right_pos score E-value substitutions power
#-----
* 98 106 121.65645 2.25295e-05 45 0.48
* 122 137 91.44593 0.000357632 57 0.58
* 96 108 88.43400 0.000466924 26 0.28
* 120 139 74.80289 0.00162024 87 0.76
* 119 140 58.72158 0.00678565 90 0.78
* 121 138 58.34837 0.00691674 99 0.82
* 94 110 57.27959 0.00760538 37 0.40
* 124 134 55.67692 0.0086606 20 0.21
* 123 135 54.59630 0.00946822 72 0.68
* 99 105 53.44797 0.0107226 15 0.14
* 97 107 44.91842 0.0405594 58 0.59

# The given structure
# SS_cons ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons ::::::::::::::::::::::::::::::::::::::::::<<<<<<____>>>>>>>><<<<<<--<<
#
# SS_cons <<<<<____>>>->>>>>-->>>>>>::
#

# Power analysis of given structure
#
# covary left_pos right_pos substitutions power
#-----
* 94 110 37 0.40
* 95 109 28 0.31
* 96 108 26 0.28
* 97 107 58 0.59
* 98 106 45 0.48
* 99 105 15 0.14
100 104 20 0.21
111 148 0 0.00
112 147 18 0.18
113 146 1 0.00
114 145 15 0.14
115 144 49 0.52
116 143 106 0.84
* 119 140 90 0.78
* 120 139 87 0.76
* 121 138 99 0.82
* 122 137 57 0.58
* 123 135 72 0.68
* 124 134 20 0.21
125 133 31 0.34

#
# BPAIRS 20
# avg substitutions per BP 43.7
# BPAIRS expected to covary 8.3
# BPAIRS observed to covary 11
#
#
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTp 0.05 [-9.78,121.66] [0 | 11 17 11 | 64.71 100.00 78.57]
#
# in_fold in_given left_pos right_pos score E-value substitutions power
#-----
* * 98 106 121.65645 2.25295e-05 45 0.48
* * 122 137 91.44593 0.000357632 57 0.58
* * 96 108 88.43400 0.000466924 26 0.28
* * 120 139 74.80289 0.00162024 87 0.76
```

```

*          *          119          140          58.72158          0.00678565          90          0.78
*          *          121          138          58.34837          0.00691674          99          0.82
*          *          94          110          57.27959          0.00760538          37          0.40
*          *          124          134          55.67692          0.0086606          20          0.21
*          *          123          135          54.59630          0.00946822          72          0.68
*          *          99          105          53.44797          0.0107226          15          0.14
*          *          97          107          44.91842          0.0405594          58          0.59

# The predicted CaCoFold structure
# SS_cons ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons ::::::::::::::::::::::::::::::::::::::::::<<<<<_____>>>>>><<<<<---<<
#
# SS_cons <<<<_____>>->>>>--->>>>:::
#

# Power analysis of CaCoFold structure
#
# covary   left_pos      right_pos      substitutions      power
#-----
*          94            110            37            0.40
          95            109            28            0.31
*          96            108            26            0.28
*          97            107            58            0.59
*          98            106            45            0.48
*          99            105            15            0.14
          111           148            0            0.00
          112           147            18            0.18
          113           146            1            0.00
          114           145            15            0.14
          115           144            49            0.52
*          119           140            90            0.78
*          120           139            87            0.76
*          121           138            99            0.82
*          122           137            57            0.58
*          123           135            72            0.68
*          124           134            20            0.21
#
# BPAIRS 17
# avg substitutions per BP 42.2
# BPAIRS expected to covary 6.9
# BPAIRS observed to covary 11

```

## Example of an RNA with pseudoknots

R-scape implements the CaCoFold folding algorithm capable of predicting pseudoknots and other non nested interactions using a cascade of dynamic programming algorithms. R-scape had adapted the program R2R to automatically include in the display all covarying interactions whether they are nested or not.

Consider the manA RNA motif. Both the proposed structure for manA RNA and the predicted CaCoFold structure have 2 pseudoknots with covariation support:

```
> bin/R-scape -s --cacofold tutorial/manA.sto
```

```

# The given structure
# SS_cons <<<<<_____>>>>>>:::[][[[,,<--
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons -----<<<<<_____>>>>>>--->>,,,( -( ( ( ( , ,
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons ,,,,,,,,,,,,,,,,,,,,,,,,,,<<<_____>>>,<<<<<_____
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::<<<<_____
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons _____>>>>>,,,<<<<<_____>>>>>,,)) ) ) ) --)
# SS_cons_1 _____>>>>>:::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons ) , , , <<<<<<-----<<<<_____>>>>
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::::

```



```

# BPAIRS expected to covary 33.2
# BPAIRS observed to covary 54

...

...

# The predicted CaCoFold structure
# SS_cons <<<<<<_____>>>>>>:[---[[[[[,,,<--
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons -----<<<<<<_____>>>>>>--->>, (--((--(((,
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons //////////////////////////////////////////////////////////////////<<<_____>>>, <<<<<<_____
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons _____>>>>>>, ,,<<<<---<_____>--->>>>>>, ))))--
# SS_cons_1 _____>>>>>>::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::::::::::
#
# SS_cons )--), <<<<<<-----<<<<--<_____>----->>>>>>
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 ::::::::::::::::::::::::::::::::::::::::::::::<<<--_____
#
# SS_cons ----->>>>>>, ,,<<<<_____>>>>>>]]]]-]::::
# SS_cons_1 ::::::::::::::::::::::::::::::::::::::::::::::::::::
# SS_cons_2 _____>>>>>>::::::::::::::::::::::::::::::::::
#

# Power analysis of CaCoFold structure
#
# covary left_pos right_pos substitutions power
# -----
* 1 43 61 0.61
* 2 42 61 0.61
* 3 41 72 0.68
* 4 40 104 0.83
* 5 39 106 0.84
* 6 38 126 0.90
45 346 82 0.74
49 344 15 0.14
* 50 343 16 0.16
* 51 342 26 0.28
* 52 341 77 0.71
* 53 340 26 0.28
* 57 107 38 0.41
* 58 106 16 0.16
* 72 101 71 0.68
* 73 100 38 0.41
* 74 99 60 0.60
* 75 98 34 0.37
* 76 97 31 0.34
77 96 27 0.30
* 109 244 37 0.40
* 112 241 41 0.44
* 113 240 51 0.53
* 115 237 61 0.61
* 116 236 46 0.49
* 117 235 62 0.62
* 118 234 49 0.52
* 150 162 52 0.54
* 151 161 47 0.50
* 152 160 36 0.39
155 223 31 0.34
* 156 222 30 0.33
157 221 28 0.31
158 220 27 0.30
* 164 206 29 0.32
* 165 205 54 0.56
* 166 204 148 0.94
* 167 203 109 0.85
* 168 202 154 0.95
* 169 201 147 0.94

```

```

      210      231      48      0.51
*    211      230      77      0.71
*    212      229      71      0.68
*    213      228      59      0.60
*    214      227      75      0.70
      218      223      59      0.60
*    246      314      62      0.62
*    247      313      92      0.79
*    248      312      141     0.93
*    249      311      90      0.78
*    250      310      99      0.82
*    251      309     105     0.84
*    252      308      37      0.40
*    265      300      61      0.61
*    266      299      44      0.47
*    267      298      42      0.45
*    268      297      36      0.39
      270      279      47      0.50
      271      278      81      0.73
*    274      329      47      0.50
*    275      328      36      0.39
*    276      327      39      0.42
      278      326      42      0.45
      317      339      13      0.12
*    318      338      37      0.40
*    319      337      60      0.60
*    320      336      18      0.18
      321      335       0      0.00
#
# BPAIRS 68
# avg substitutions per BP 58.0
# BPAIRS expected to covary 36.2
# BPAIRS observed to covary 55

```

The “SS\_cons\_1” and “SS\_cons\_2” lines describe the interactions that are not nested relative to the main “SS\_cons” structure.

R-scape uses R2R to produce figures of the consensus structures where pseudoknots are also annotated. R-scape] option -s produces the file **tutorial/manA.R2R.sto.{pdf,svg}** with the structure annotated in the input alignment. R-scape] option -cacofold produces the file **tutorial/manA.fold.R2R.sto.{pdf,svg}** with the structure produced by R-scape. See Figure 1.

## Single sequence structure prediction

If the alignment includes only one sequence, no statistical test is performed.

```
> bin/R-scape --cacofold tutorial/manA-oneseq.sto
```

reports the best structure given the sequence. No covariation support is possible for any of the basepairs reported from this analysis. Structures produced this way have to taken with great skepticism.

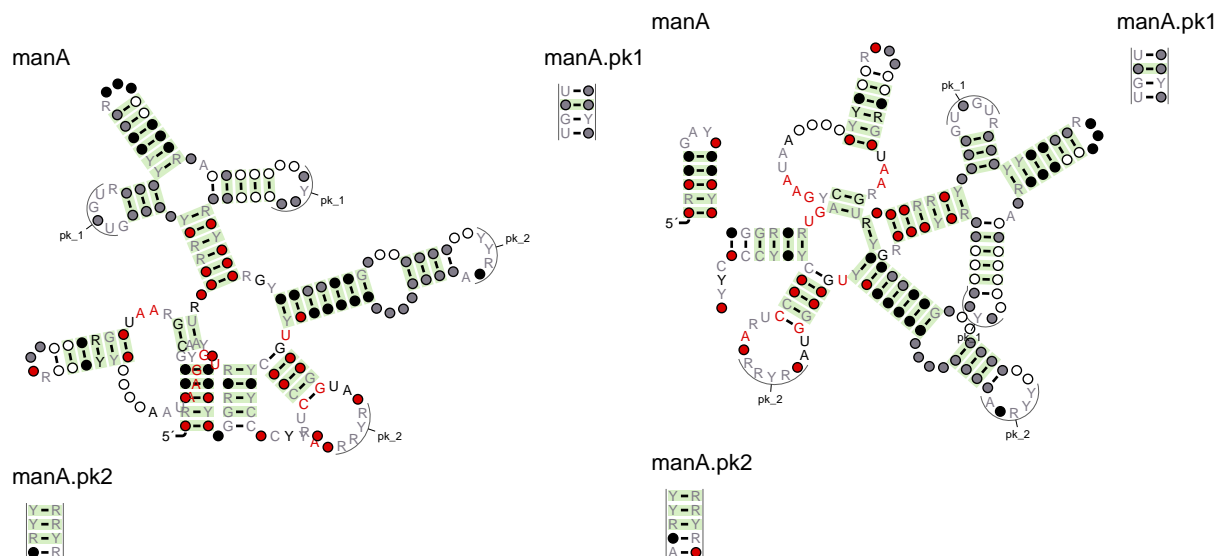


Figure 1: **Left:** `tutorial/manA.R2R.sto`.`{pdf,svg}`, the consensus secondary structure given in the input alignment, depicted by R-scape, using the program R2R. **Right:** `tutorial/manA.fold.R2R.sto`.`{pdf,svg}`, The consensus structure produced by R-scape (option `-cacofold`). Base pairs with covariation scores equal or below the target E-value (0.05 as default) are depicted in green.

## 5 Tutorial for running CaCoFold-R3D

Here's a tutorial walk-through of how to use CaCoFold-R3D. CaCoFold-R3D, extends CaCoFold by reporting a predicted structure that incorporates standard Watson-Crick helices together with 3D RNA motifs that are composed of non-Watson-Crick pairs and unpaired residues. As with CaCoFold, the CaCoFold-R3D structure may contain pseudoknots and triplets, provided that they present covariation signal.

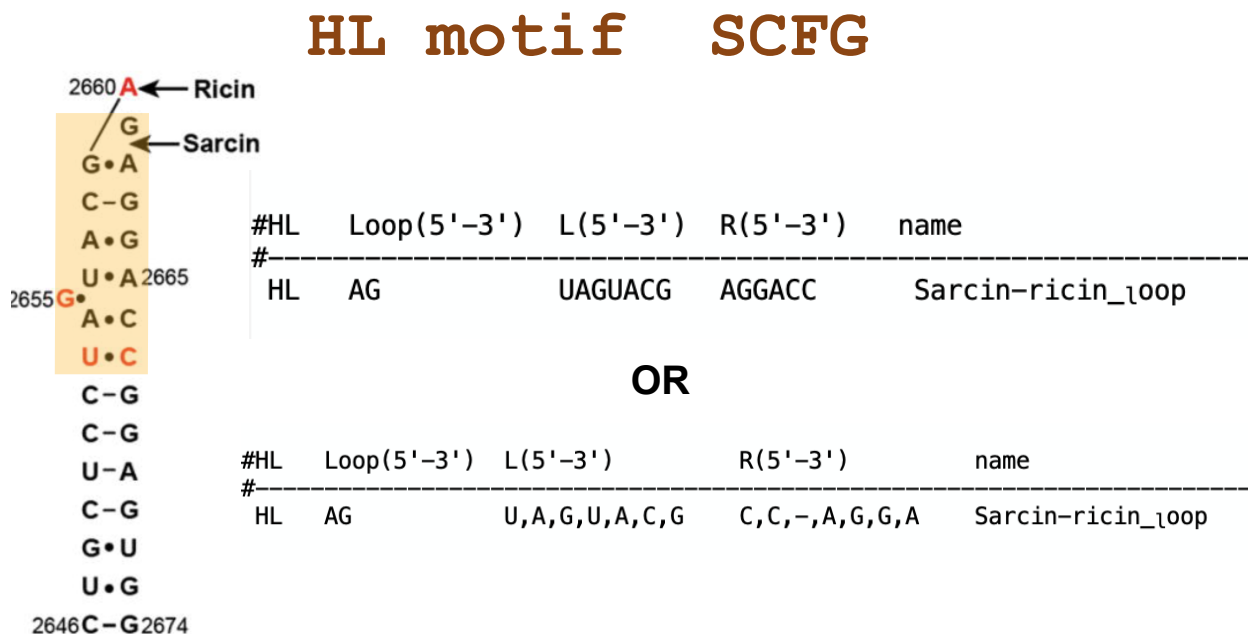
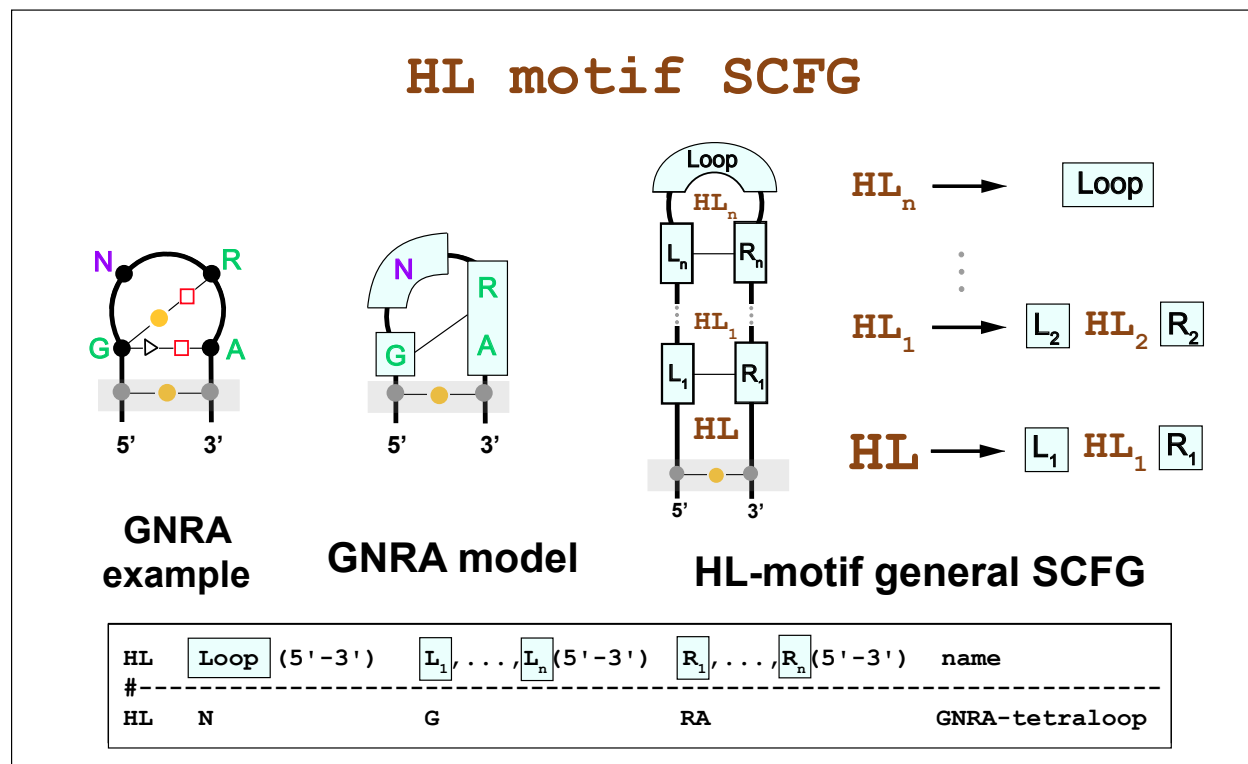
### The `-cacofold -r3d` options

After performing one of the two statistical tests with R-scape, these two options together implements the CaCoFold-R3D algorithm.

## Examples of 3D motifs and how to incorporate them to a R3D descriptor file

### Hairpin Loops (HL)

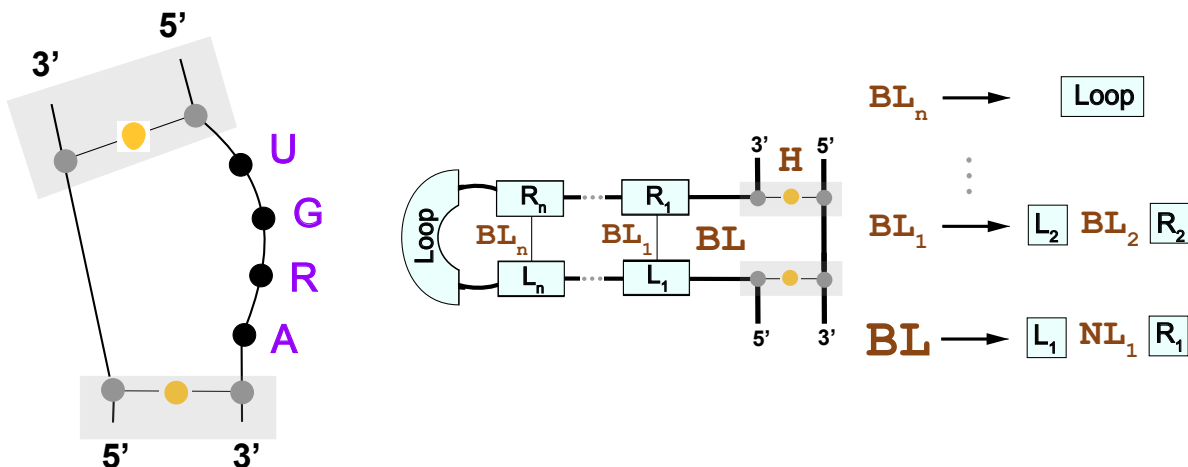
Here we present the general HL SCFG model, and two example of HL motifs with their corresponding descriptor.



## Bulge Loops (BL)

Here we present the general BL SCFG model, and two example of BL motifs with their corresponding descriptor.

### BL motif SCFG

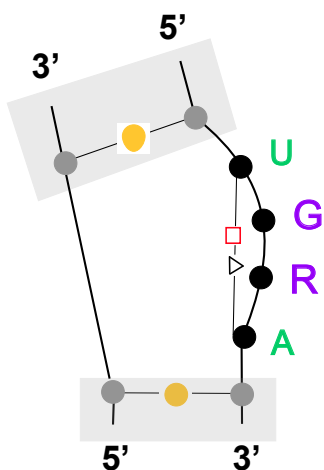


## Docking elbow model

## BL-motif general SCFG

#BL	Loop(5'-3')	L(5'-3')	R(5'-3')	name
#				
BL	UGRAA	-	-	Docking-elbow

### BL motif SCFG



#BL	Loop(5'-3')	L(5'-3')	R(5'-3')	name
#				
BL	GR	U	A	Fake-bulge

Here we present the general IL SCFG model, and two example of IL mofits with their corresponding descriptor.

**K-turn example**

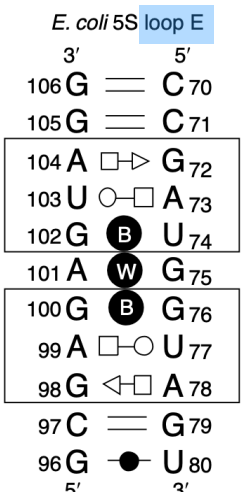
**K-turn model**

**IL-motif general SCFG**

#IL	Loop-L(5'-3')	Loop-R(5'-3')	L-o(5'-3')	R-o(5'-3')	L-i(5'-3')	R-i(5'-3')	name
#							
IL	-	RNN	G,A	A,R	-	-	K-turn

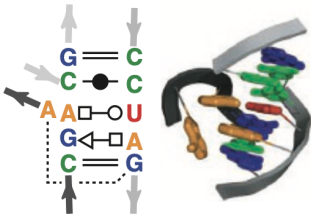
IL motif SCFG

Loop-E



#IL	Loop-L(5'-3')	Loop-R(5'-3')	L-o(5'-3')	R-o(5'-3')	L-i(5'-3')	R-i(5'-3')	name
#	IL	GAG	UGG	G,A	U,A	U,A	A,G
							Loop-E

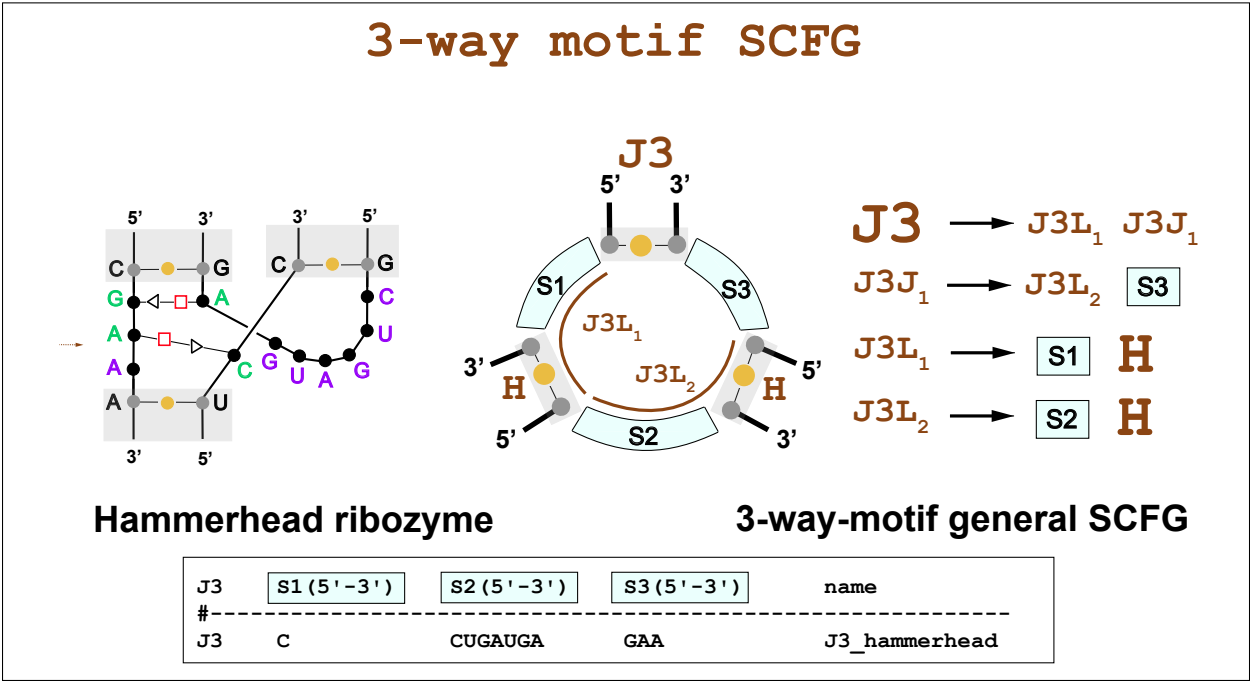
hook-turn



#IL	Loop-L(5'-3')	Loop-R(5'-3')	L-o(5'-3')	R-o(5'-3')	L-i(5'-3')	R-i(5'-3')	name	
#	IL	A	-	G,A	A,U	C	C	hook-turn

3-way Junctions (J3)

Here we present the general J3 SCFG model, and one example of J3 motifs with their corresponding descriptor.



Here we present the general J4 SCFG model, and one example of J4 mofits with their corresponding descriptor.

Example of a 4-way Junction.

**HCV IRES**

**4-way-motif general SCFG**

Transitions:

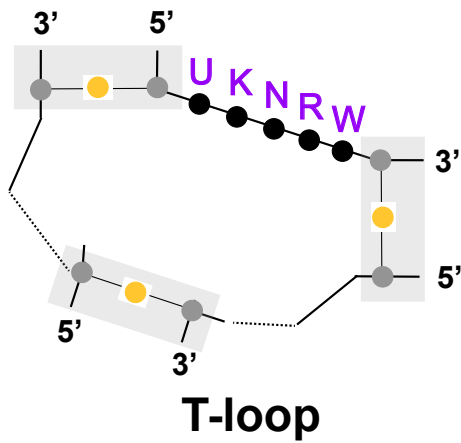
- $J4 \rightarrow J4L_1 \ J4J_1$
- $J4J_1 \rightarrow J4L_2 \ J4J_2$
- $J4J_2 \rightarrow J4L_3 \ S4$
- $J4L_1 \rightarrow S1 \ H$
- $J4L_2 \rightarrow S2 \ H$
- $J4L_3 \rightarrow S3 \ H$

#J4	S1(5'-3')	S2(5'-3')	S3(5'-3')	S4(5'-3')	name
J4	-	AA	-	U	J4_HCV_IRES

## Branch Segments (BS)

BS sequence segments can appear in any higher order junction. Here we provide the general form of a BS SCFG, and the particular case of a T-loop segment.

### BS motif SCFG



BS → Loop

### BS-motif general SCFG

#BS	Loop(5'-3')	name
#		
BS	UKNRW	T-loop

## The -r3dfile to customize the descriptor file

CaCoFold-R3D reads a descriptor file to identify all the 3D motifs, and to build the probabilistic model incorporating all of them. The default descriptor file used is given below (lines starting with # are comments).

Using the option `--r3dfile`, one can introduce any customized combination of 3D motifs to consider.

```
#HL Loop(5'-3') L(5'-3') R(5'-3') name
#-----
HL N G RA GNRA-tetraloop
HL URA - - U-turn
HL UNCG - - UNCG-tetraloop
HL ANYA - - ANYA-tetraloop
HL CUYG - - CUYG-tetraloop
HL YGNN - - YGNN-tetraloop
HL GANC - - GANC-tetraloop
HL UNAC - - UNAC-tetraloop
HL URRR - - T-loop-tetraloop
HL UAACR - - L8_RNaseP_bact_a
HL AG UAGUACG AGACC Sarcin-ricin_loop
HL AGGAY - - CsrA_binding
HL GAGUA - - GAGUA_pentaloop
HL AA - - AA-5SrRNA
HL GCRYA - - U6-loop
#BL Loop(5'-3') L(5'-3') R(5'-3') name
#-----
BL UGRAA - - Docking-elbow
BL AA - - AA-bulge
BL G - - bulged-G
#IL Loop-L(5'-3') Loop-R(5'-3') L-o(5'-3') R-o(5'-3') L-i(5'-3') R-i(5'-3') name
#-----
IL RNN - G,A A,R - - K-turn
IL - RNN G,A A,R - - K-turn-b
IL N - R,A,R,R A,U,R,R - - Loop-E
IL UAAG UAU C,C G,G - - GAAA_Tetraloop-receptor
IL CVC V - - C-loop
IL G - G,A A,A U,A A,G G-bulge
IL G - U,A C,C U,A A,G G-bulge_Das
IL - G,A A,G - - Tandem-GA
IL YCC AAC - - Twisted-up
IL YAA RAN - - UAA_GAN # 23S rRNA, RNase P
IL RRRA YYNR - - J4a/4b # riboswitch
IL AA AA - - J4/5-IL # ribozyme
IL GUA GG - - GUA^GG_RRE
IL CAGG AGCA - - S_domain
IL GAAC CUAG - - Hook-turn
IL UGRAA - - Docking-elbow
IL UU AUU - - pK-turn
#J3 S1(5'-3') S2(5'-3') S3(5'-3') name
#-----
J3 N CUGA A J3_hammerhead
J3 U YUCUAC AC J3_purine
J3 NN NNNNNN NN J3_typeA
J3 NNNN NNNN NNNN J3_typeB
J3 NN NNN NNNNNN J3_typeC
J3 - UGAGA - J3_TPP #Breaker
#J4 S1(5'-3') S2(5'-3') S3(5'-3') S4(5'-3') name
#-----
J4 - AA - U J4_HCV_IRES
J4 N - NNN - J4_tRNA
J4 N N NN - J4_manA
J4 R - - R J4_U1RNA
#BS Loop(5'-3') name
#-----
BS UKNRW T-loop
BS RRGU LoopE-a
BS RARR LoopE-b
BS AAYAARAACAANARR CRC_binding
```

## 6 Inputs

### The Stockholm format to describe a consensus structure

The input is a multiple sequence alignment in Stockholm format ([https://en.wikipedia.org/wiki/Stockholm\\_format](https://en.wikipedia.org/wiki/Stockholm_format)). The Stockholm format allows to provide a consensus structure for the alignment using the tag `#=GC SS_cons`. R-scape then can analyze the given consensus structure using option `-s`.

The Stockholm format uses symbols `()`, `<>`, `{}`, and `[]` to describe a nested structure. It also allows non-nested structures such as pseudoknots using the symbols `Aa` or `Bb`, ..., `Zz`. See file `tutorial/RF00162.SAM.pk.sto`, where a pseudoknot with 4 basepairs is annotated for the SAM-I riboswitch.

### A extended Stockholm format to describe consensus structures with overlapping base pairs

The `#=GC SS_cons` annotation does not allow to display other pairs that overlap with the rest of the structure. These overlapping pairs can be base triplets, other non Watson-Crick interactions or even alternative structures. CaCoFold identifies a number of these overlapping pairs with covariation support. In order to annotate these overlapping basepairs, R-scape uses additional annotations in the form `#=GC SS_cons_1`, `#=GC SS_cons_2`,... See for example file `tutorial/RF00162.SAM.CaCoFold.sto`, where in addition to the pseudoknot there are two other overlapping pairs annotated as part of the structure.

You should use this augmented Stockholm format to input a consensus structure with overlapping basepairs. If you run

```
bin/R-scape -s tutorial/RF00162.SAM.CaCoFold.sto
```

you will see that the whole structure is taken into account in the statistical test.

## 7 Outputs

A Stockholm alignment file can include several different multiple sequence alignments (MSAs). For each alignment file `rnafile.sto`, R-scape produces the following output files, one for each individual alignment in an input Stockholm file:

- `rnafile.msaname.cov`** Tabular output with the significant pairs, with their score and E-value, estimated number of substitutions and power.
- `rnafile.msaname.sorted.cov`** Tabular output sorted from highest to lowest E-value.
- `rnafile.msaname.power`** Tabular output with the list of basepairs in the proposed RNA structure annotated their power. The file also reports the alignment power, and the expected number of basepairs to covary.

### Covariation tabular output

The distribution includes in the directory `tutorials/` examples of output files. If you run R-scape, the outputs will go into your current working directory (not necessarily `tutorials/`).

The output file `tutorial/updated_Arisong.1.cov` looks like this:

```
> more tutorial/updated_Arisong.1.cov

# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTp      0.05      [-9.78,121.66]    [0 | 11 20 11 | 55.00 100.00 70.97]
#
# in_given left_pos right_pos score E-value substitutions power
#-----
*           94      110    57.27959 0.00760538      37      0.40
*           96      108    88.43400 0.000466924     26      0.28
...
```

The output file is a tabular list of significant pairs sorted by sequence positions:

**First column** indicates whether the significant pair is part of the given structure (\*), or not. If the pair is not in the structure, we distinguish whether the pair is compatible with the given structure (~) or not (blank).

In addition, if the structure is provided by a PDB file (using the option `--pdb`), a non Watson-Crick/Watson-Crick base pair is designated by “\*\*\*”. A contact that is not a basepair is designated by: “*c* ~” if compatible with all the basepairs, or by “*c*” otherwise.

**Second and third columns** are the two positions of the pair,  $i \leq j$  respectively. Positions are relative to the input alignment.

**Fourth column** is the covariation score.

**Fifth column** is the E-value. Significant positions have E-values  $\ll 1$ .

**Sixth column** is the estimated number of total substitutions in the two columns.

**Seventh column** is the basepair power or probability that it should covary.

The output file also includes two comment lines per alignment in the file:

**First comment line** describes properties of the alignment: number of sequence (nseq), alignment length (alen), average percentage identity (avgid), and number of base pairs (nbpairs). Values in parentheses correspond to the alignment as given. Values not in parentheses correspond to the analyzed alignment after the filters (for redundant sequences and gapped columns) have been applied.

Second comment line describes properties of the R-scape search: the covariation method (GTp), the E-value threshold (0.05), the range of scores for all pairs in the alignments (from -9.7 to 89.1), the number of covarying non base pairs (0), the number of covarying base pairs (11), the number of base pairs (20), and the total number of covarying pairs (11). Lastly we provide the sensitivity (SEN=55.00=11/20), positive predictive value (PPV=100.00=11/11), and F-measure ( $F=70.97 = 2 * SEN * PPV / (SEN+PPV)$ ).

## Power tabular output

The output file `tutorial/updated.Arisonong.1.power` looks like this:

```
> more tutorial/updated.Arisonong.1.power

# Power analysis of given structure
#
# covary left_pos right_pos substitutions power
#-----
#
#   *   94       110       37       0.40
#       95       109       28       0.31
#   *   96       108       26       0.28
#   *   97       107       58       0.59
#   *   98       106       45       0.48
#   *   99       105       15       0.14
#       100      104       20       0.21
#
#   .
#   .
#   .
#   *   122      137       57       0.58
#   *   123      135       72       0.68
#   *   124      134       20       0.21
#       125      133       31       0.34
#
#
# BPAIRS 20
# avg substitutions per BP 43.7
# BPAIRS expected to covary 8.3
# BPAIRS observed to covary 11
```

This file includes the list of all basepairs in the proposed structure given with the input alignment. Each basepair is annotated with the estimated number of substitutions and power.

## Default graphical outputs

By default, the following files are also produced

- `rnafilename.msaname.R2R.sto`** Stockholm file annotated by a modified version of the R2R program. This file includes the information necessary to draw the consensus structure, and to annotate the significantly covarying base pairs.
- `rnafilename.msaname.R2R.sto.{pdf,svg}`** Drawing of the R-scape-annotated consensus secondary structure.
- `rnafilename.msaname.surv`** A two column file with the survival functions (surv) for the covariation scores.
- `rnafilename.msaname.surv.ps`** Plot of the score's survival function  $P(X > \text{score})$ . Drawing this file requires that program **gnuplot** is installed somewhere in the  $\$ \{ \text{PATH} \}$ , or that the environmental variable **GNUPLOT** pointing to a gnuplot executable is defined.
- `rnafilename.msaname.dplot.{ps,svg}`** Dot plot of the consensus secondary structure annotated according to covariation. Drawing of this file requires that program **gnuplot** is installed somewhere in the  $\$ \{ \text{PATH} \}$ , or that the environmental variable **GNUPLOT** pointing to a gnuplot executable is defined.

For each alignment, **msaname** is given by <ACC>\_<ID>, the combination of the accession #=**GF AC** <ACC> and name #=**GF ID** <ID> in the Stockholm-format markups (or one of two if the other is not defined). If none of those fields are defined, **msaname** is a number describing the order in the file of the given alignment.

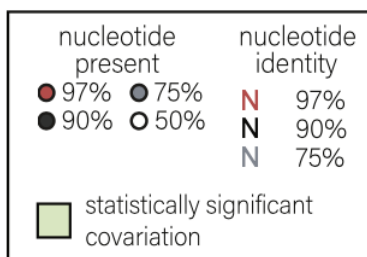
## Details about graphical outputs

Two files are produced per alignment in the input file:

File **tutorial/updated.Arisong.1.R2R.sto** is a Stockholm formatted alignment that includes the input alignment annotated with the consensus structure. This Stockholm file also includes the additional annotation required to use the drawing program R2R.

It is possible that the resulting drawing will show parts of the secondary structure occluded from each other (especially for long RNAs). Using this file, one can customize a different drawing of the structure using the R2R documentation, provided in `lib/R2R/R2R-manual.pdf`.

File **tutorial/updated.Arisong.1.R2R.sto.pdf** depicts the consensus structure with the R-scape covariation annotation using the R2R software to depict the alignment. The defaults used by R-scape to depict the alignment positions are given in this legend,



File **tutorial/updated.Arisong.1.surv** looks like this:

```
> more tutorial/updated.Arisong.surv
```

```
121.795428      0.05
95.862635      0.1
89.113004      0.15
...
&
63.890698      0.000485437
58.917286      0.000970874
47.904730      0.00145631
...
&
81.652885      2.40385e-06
77.745204      4.80769e-06
77.034717      7.21154e-06
...
&
256.788050      2.64342e-17
256.432807      2.7899e-17
256.077563      2.94449e-17
...
&
```

The first column is a covariation score ( $x$ ). The second column is the survival function  $P(X > x)$ , that is the frequency of pairs having score larger than  $x$ . The file includes four survival functions separated by a “&” line. The three survival functions correspond to:

First functions: the given alignment, proposed base pairs. (This section is empty if no secondary structure is proposed.)

Second functions: the given alignment, not proposed pairs.

Third function: the aggregation of all null alignments, all possible pairs.

Fourth function: the expected null survival function according to the tail Gamma fit.

## Using option `--cacofold`

If the option `--cacofold` is used, R-scape produces the following additional files describing the maximal-covariation optimal secondary structure:

`rnafilename.fold.sto` The original alignment with the R-scape structural annotation

`rnafilename.fold.R2R.sto` File used by R2R to display the R-scape structure

`rnafilename.fold.R2R.sto.{pdf,svg}`

`rnafilename.fold.surv`

`rnafilename.fold.surv.{ps,svg}`

`rnafilename.fold.dplot.{ps,svg}`

These files are formatted identically to those describing the given consensus structure.

## Graphical outputs per alignment

Three plots are produced per alignment in the input file:

updated\_Arisong\_1

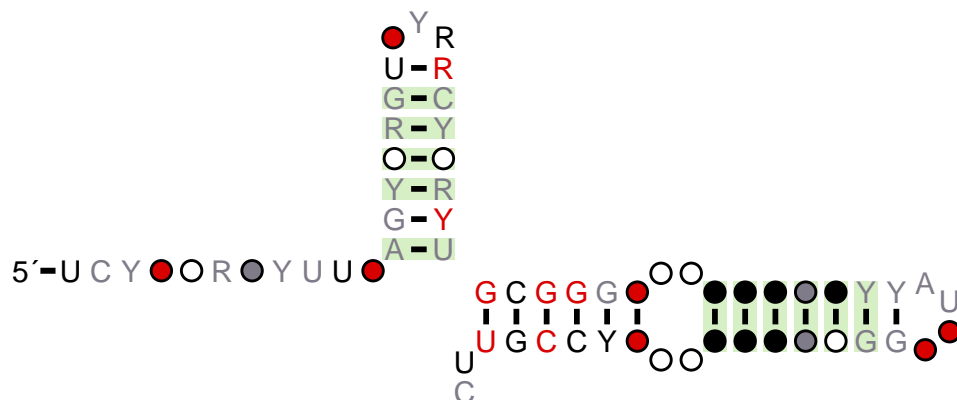


Figure 2: **tutorial/updated\_Arisong.1.R2R.sto.{pdf,svg}**: annotated consensus secondary structure. Base pairs with covariation scores equal or below the target E-value (0.05 as default) are depicted in green. By default only positions in the alignment with more than 50% occupancy are depicted (unless they form a base pair). Option `--r2rall` forces the depiction of all positions in the alignment.

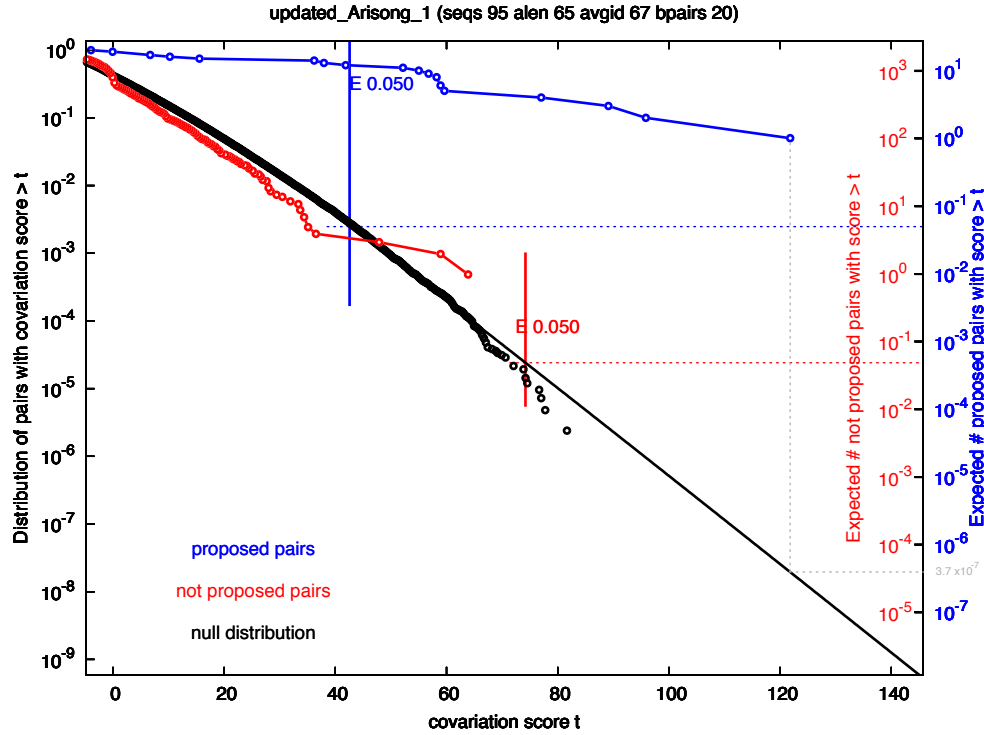


Figure 3: `tutorial/updated_Arisong_1.surv.{ps,svg}`: covariation scores survival function  $P(X > x)$ . The survival function of scores for all pairs in the given alignment is depicted in blue. The survival function for the null alignments is depicted in black. A black line indicates to fit to a truncated Gamma distribution of the tail of the null distribution. In red, we plot the survival function of scores for the pairs in the given alignment excluding those proposed as base pairs. For a particular pair, as an example the highest scoring one from the distribution of proposed pairs (blue), we obtain its E-value by drawing a vertical (gray) line from the point to the null distribution (black). The corresponding value in the blue scale gives us the E-value for that pair (in this example,  $3.7 \cdot 10^{-7}$ ).

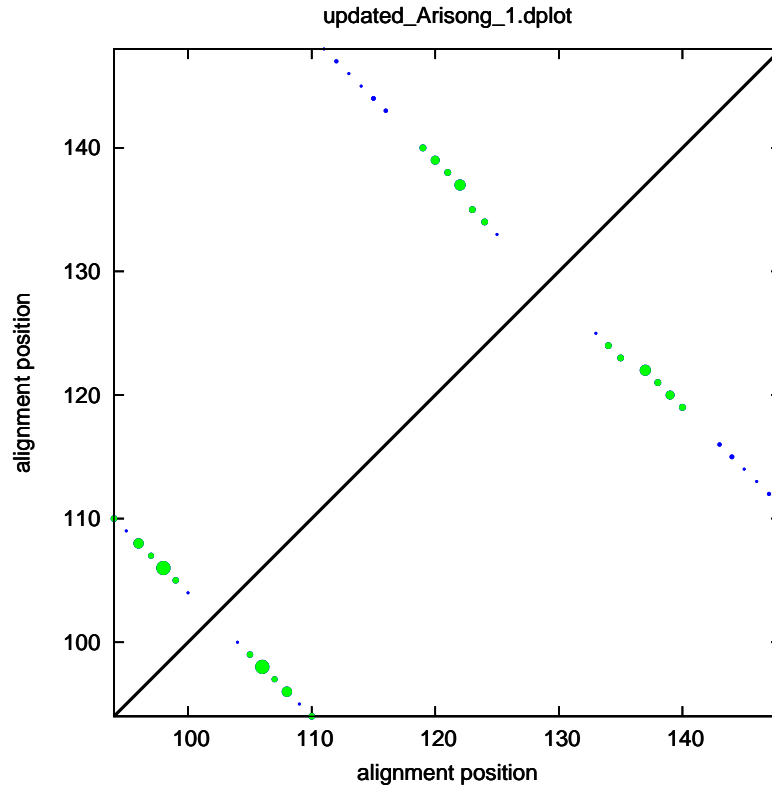


Figure 4: `tutorial/updated_Arisong_1.dplot`. `{ps,svg}:dotplot`. Dot size is proportional to the covariance score. In blue we depict the consensus base pairs; in green, the consensus base pairs that show significant covariation; in orange (none shown in this plot), we depict other pairs that have significant covariation, are not part of the consensus secondary structure but are compatible with it; in black we depict other significant pairs. Position are relative to the original input alignment (before any gapped column is removed).

## 8 Options

The whole list of options can be found using

```
> R-scape -h
```

Some important options are:

### Covariation statistic options

**-E <x>**

Target E-value is  $x \geq 0$ .

**--GT, --MI, --MIr, --MIg, --CHI, --OMES, --RAF, --RAFS,**

We favor the G-test covariation statistic, but a total of eight covariation statistics are currently implemented in R-scape. For each covariation statistic (GT, for instance), R-scape can also calculate its average product correction (GTp) and its average sum corrections (GTa). For each option above, appending “p” or “a” chooses one of the corrections. For example, --GT does the G-test statistic, --GTp does the APC-corrected G-test statistic, --GTa does the ASC-corrected G-test statistic.

The R-scape default is --GTp.

Details of the definition and provenance of the different covariation statistics can be found in the R-scape manuscript: Rivas, E. & Eddy S. E., “A statistical test for conserved RNA structure shows lack of evidence for structure in lncRNAs”.

In a nutshell, given two alignment columns  $i, j$ ,

G-test:(Woelf, 1957)	$GT(i, j) = 2 \sum_{a,b} \text{Obs}_{ij}^{ab} \log \frac{\text{Obs}_{ij}^{ab}}{\text{Exp}_{ij}^{ab}},$
Pearson’s chi-square:	$CHI(i, j) = \sum_{a,b} \frac{(\text{Obs}_{ij}^{ab} - \text{Exp}_{ij}^{ab})^2}{\text{Exp}_{ij}^{ab}},$
Mutual information:(Shannon, 1948; Gutell et al., 1994)	$MI(i, j) = \sum_{a,b} P_{ij}^{ab} \log \frac{P_{ij}^{ab}}{p_i^a p_j^b},$
MI normalized:(Martin et al., 2005)	$MIr(i, j) = \frac{MI(i, j)}{H(i, j)} = \frac{MI(i, j)}{-\sum_{a,b} P_{ij}^{ab} \log P_{ij}^{ab}},$
MI with gap penalty:(Lindgreen et al., 2006)	$MIg(i, j) = MI(i, j) - \frac{N_{ij}^G}{N},$
Obs-Minus-Exp-Squared:(Fodor and Aldrich, 2004)	$OMES(i, j) = \sum_{a,b} \frac{(\text{Obs}_{ij}^{ab} - \text{Exp}_{ij}^{ab})^2}{N_{ij}},$
RNAalifold (RAF):(Hofacker et al., 2002)	$RAF(i, j) = B_{i,j},$
RNAalifold Stacking (RAFS):(Lindgreen et al., 2006)	$RAFS(i, j) = \frac{1}{4} (B_{i-1,j+1} + 2 B_{i,j} + B_{i+1,j-1}).$

where  $a, b$  are (non-gap) residues;  $N$  is the total number of aligned sequences;  $\text{Obs}_{ij}^{ab}$  is the observed count of  $a : b$  pairs in columns  $i, j$  (only counting when both  $a, b$  are residues);  $N_{ij}$  is the total number of residue pairs in columns  $i, j$  (only counting when both  $a, b$  are residues);  $P_{ij}^{ab}$  is the observed frequency of pair  $a : b$  in columns  $i, j$  ( $P_{ij}^{ab} = \frac{\text{Obs}_{ij}^{ab}}{N_{ij}}$ );  $\text{Exp}_{ij}^{ab} = N_{ij} p_i^a p_j^b$  is the expected frequency of pair  $a : b$  assuming  $i, j$  are independent, where  $p_i^a$  are the marginal frequencies of  $a$  residues in column  $i$  (averaged to all other positions) ( $p_i^a = \frac{1}{L-1} \sum_{j \neq i} \sum_b P_{ij}^{ab}$ );  $N_{ij}^G = N - N_{ij}$  is the number of pairs involving at least one gap symbol; the definition of  $B_{i,j}$  used in the RAF and RAFS statistics is involved, a concise definition can be found elsewhere (Lindgreen et al., 2006).

The background corrections (Dunn et al., 2007) for a given covariation statistic above  $\text{COV}(i, j)$  are,

$$\begin{aligned} \text{Average product correction} \quad \text{COVp}(i, j) &= \text{COV}(i, j) - \frac{\text{COV}(i) \text{COV}(j)}{\text{COV}}, \\ \text{Average sum correction} \quad \text{COVa}(i, j) &= \text{COV}(i, j) - (\text{COV}(i) + \text{COV}(j) - \text{COV}). \end{aligned}$$

**--C2, --C16, --CWC**

For all the covariation statistics (except RAF and RAFS), one can do a 16-component (C16) or a two-component (C2) calculation, depending on whether it uses the 16 possible pair combinations, or those are group in two classes depending on whether they form a Watson-Crick pair (6 cases, including U:G and G:U), or whether they do not (10 cases).

R-scape's default is the 16 component covariation statistic, unless the number of sequences in the alignment is  $\leq 8$  or the length of the alignment is  $\leq 50$ , in which case it uses the two-class covariation statistic.

Option CWC (implemented for GT, GTp and GTa) uses a 6 component covariation statistic which only considers Watson-Crick-like pairs: A-U, U-A, C-G, G-C, G-U, U-G.

## Options to calculate power

**--singlesubs**

Default option. The base pair substitutions are calculated as the sum of the individual substitutions observed for each of the positions in the base pair.

**--joinsubs**

The base pair substitutions are calculated as the sum of the individual substitution observed for each positions in the base pair, but using only sequences in which both positions are occupied, that is, cases where one of the two positions is a gap are ignored.

**--doublesubs**

The base pair substitutions are calculated a the subset of substations in which both residues have changed.

## Covariation aggregation options

**--fisher, --sidak**

Two different options to produce aggregated E-values calculated for each helix in the proposed or CaCoFold structure.

**--lancaster, --wfisher**

Two different options to produce aggregated E-values calculated for each helix in the proposed or CaCoFold structure.

The Lancaster aggregation uses the number of substitutions per base pair. The weighted fisher (wfisher) aggregation uses the power per base pair. These two options require the use of the default option **--singlesubs** in order to calculate the number of substitutions and power for each base pair respectively.

**--lancaster\_join, --wfisher\_join**

These two options require the use of the option **--joinsubs** in order to calculate the number of substitutions and power for each base pair respectively.

**--lancaster\_double, --wfisher\_double**

These two options require the use of the option **--doublesubs** in order to calculate the number of substitutions and power for each base pair respectively.

## Search options

### **-s**

The “two-set test” option. This option requires that a structure is provided with the alignment. If option `-s` is used, R-scape performs two independent test, one for the given structure, another for all other possible pairs. The default is a “one-set test” in which all possible pairs in the alignment are tested equivalently.

### **--cacofold**

A CaCoFold structure is computed that includes all significant base pairs. All files related to this CaCoFold structure include the suffix `.cacofold`.

When option `--cacofold` is used, a file with the original alignment annotated with the R-scape structure in Stockholm format is produced. This alignment has the suffix `.cacofold.sto`.

### **--naive**

Reports the laundry list of all covariation scores, without any statistical significance (E-value) associated to them. No null alignments are created.

### **--tstart <n>**

Analyze starting from position  $n \geq 1$  in the alignment.

### **--tend <n>**

Analyze ending at position  $n \leq L$  in the alignment.

### **--window <n>**

R-scape can be run in a window scanning version for long alignments. The window size is  $n > 0$ .

### **--slide <n>**

In scanning mode, this options sets the number of positions to move from window to window,  $n > 0$ .

### **--vshuffle**

Vertical shuffle, a developers tool. Before performing any analysis, it shuffles all residues in each alignment column independently.

### **--cshuffle**

Column shuffle, a developers tool. Before performing any analysis, it shuffles all columns in the alignment.

### **--givennull <f>**

Use histogram provided in file `<f>` as null.

## Input alignment options

### **-I <x>**

Only sequences with less than  $0 < x \leq 1$  pairwise similarity are considered in the analysis. Pairwise % identity is defined as the ratio of identical positions divided by the minimum length of the two sequences. If this option is not used all (weighted) sequences are used in the analysis.

### **--gapthresh <x>**

Only columns with less than  $0 < x \leq 1$  fraction of gaps are considered in the analysis.

### **--consensus**

If the alignment has a GC “seq.cons” field, only consensus positions will be analyzed.

### **--submsa <n>**

Analyzes a random subset of the input alignment.

### **--treefile <f>**

A phylogenetic tree in Newick format can be given (by default a tree is created from the alignment using the program FastTree (v2.1.11) (Price et al., 2010)). R-scape checks that the number of taxa and the names of the taxa matches for all alignments analyzed.

### **--ntree <n>**

Number of different FastTree trees to use when generating the null alignments. Default is one tree, the one resulting from feeding the input alignment to FastTree. if `--ntree > 1`, the rest of the trees are generated after randomly rearranging the sequences in the alignment.

Because FastTree is not deterministic, altering the order of the sequences in the alignment can result in slightly different trees, and in some rare occasions that can results in different distribution of null covariation scores. In those cases, it is recommended to generate null alignments from different trees obtained from randomly rearrange the sequences in the alignment.

Option `--ntree <n>` is incompatible with option `--treefile <f>` which inputs a particular tree.

## Options for producing a CaCoFold structure

### **--cacofold**

When using the option `--cacofold`, R-scape engages the CaCoFold algorithm to produce a predicted structure. The CaCoFold algorithm incorporates all positive (significantly covarying) base pairs, and prevents any negative pair (pairs that have power of covariation but not covariation) from happening. The CaCoFold algorithm uses a recursive cascade of constrained foldings. The first fold uses the RBG probabilistic grammar, the rest use the G6X probabilistic grammar. To find secondary structure and RNA 3D motifs:

### **--cacofold --r3d**

to find the secondary structure and RNA 3D motifs.

### **--r3dfile <f>**

provide a customized r3d file. The defaults file is at `data/r3d/R3D.grm`  
Regarding the predicted structure, CaCoFold can use one two algorithms:

#### **--cyk**

Default option. Each folding reports the structure with the best probability using the CYK algorithm.

#### **--decoding**

This options returns the structure obtained by posterior decoding.

Posterior decoding usually performs better than CYK. Both algorithm has the same algorithmic complexity. CYK is faster.

Several additional options can be used in combination with `--cacofold`,

#### **--refseq**

By default the CaCoFold algorithm folds a profile sequence built from the alignment. Using this option, the sequence to fold is a consensus reference sequence.

#### **--E\_neg <x>**

Pairs with E-value larger than the E-value cutoff but smaller than  $|x|_L$  will not be called negatives regardless of their covariation power. Default for E\_neg is 1.0.

#### **--lastfold**

This option forces one last alternative fold (using grammar G6X) after all covarying base pairs have already been integrated into the structure. By default this last fold is not performed. In the absence of any covarying base pair, one fold is performed using grammar RBG.

#### **--show\_hoverlap**

This option leaves the alternative helices unmodified. By default, alternative structures are trimmed down to show no overlap with helices from the previous layers.

#### **--covmin <n>**

Minimum distance between position to report significant covariation. Default is 1, which means that significant covariation between contiguous positions are reported.

#### **--allow\_negatives**

This option (just for developers) allows all base pairs to form regardless of their power.

#### **--Rfam**

This option is meant to be used by the Rfam curators when using CaCoFold to propose improved consensus structures for an Rfam family. It removes covariation that cannot be taking into account by the Rfam models. Thus, maybe missing important covariation information that is not compatible with RNA secondary structure.

#### **--rmcoding**

This option removes from the R2R display the covariation that appears to be due to protein-coding codon signals instead of RNA structure. This affect only to the display figures not to anything in the analysis of covariation.

Using options `--cacofold --Rfam`, the CaCoFold structure is trimmed such that:

- Base pairs have to have at least 3 nucleotides of separation (covarying pairs removed if they don't).

- Overlaps between helices are trimmed down if possible without removing any covarying pair.
- Pseudoknots (pk) are kept, but alternative motifs identified as: triplets (tr), cross (xcov), or side (scov) covariations are removed.
- Base pairs which appear to be non WC (defined by the observed frequency in the alignment of the pair being A:U, U:A, C:G, G:C, G:U or U:G being less than 0.3) are removed, even if they covary.

## Options for importing a structure

R-scape does not require to input a structure (either a RNA structure or a protein contact map). By default R-scape analyzes all possible pairs in the alignment.

There are two ways to provide a contact map (or structure):

- By providing the alignment in Stockholm format with a “ss\_cons” field including the consensus structure for the alignment. (For RNA alignments only.)
- By analyzing a 3D structure provided in a PDB file. (For either RNA or peptide alignments.)

These two methods can be combined together. For a nucleotide alignment, if both a consensus structure is present in the alignment, and a PDB file is provided (using option `--pdb`), the consensus structure will be extended by the information provided by the pdbfile. To ignore the consensus structure use option `--onlypdb`.

From the PDB file we obtain three types of structural pairs:

- **Contacts:** defined as those two residues at a close spatial distance (specified by the user with option `--cntmaxD`).
- **Basepair:** RNA base pairs.  
RNA Basepair are calculated using the program `rnaview` (Yang et al., 2003).  
These RNA base pairs can be further classified in two types:
  - **Watson-Crick base pairs:** the canonical RNA base pairs. mostly A:U, G:C, or G:U pairs. (H-bond interactions between two W-C faces in cis).
  - **Other base pairs:** the non-canonical RNA base pairs (all other types of H-bond interactions, 12 different types).

Contacts and RNA base pairs are extracted as follows:

- The spatial distance between any two residues is calculated as the minimal Euclidean distance between any two atoms (excluding H atoms). Any two pairs at a distance not larger than a maximum value (`cntmaxD`) are called a “contact”.
- RNA base pairs are obtained using the program `rnaview` (Yang et al., 2003) (<http://ndbserver.rutgers.edu/ndbmodule/services/download/rnaview.html>).  
The RNA base pair annotation takes precedent over the annotation as “contact”.

The options that control the input of a structure or contact map are:

`--pdb <s>`

Reads a pdbfile associated to the alignment, and extracts the contacts from it.

A “cmap” file is produced reporting the structure obtained from the PDB file.

Option `--pdb` is incompatible with `--cacofold`.

**--cntmaxD <x>**

Maximum distance (in Angstroms) allowed between two residues to define a “contact” is  $\langle x \rangle$ .

**--cntmind <n>**

Minimum distance (in residue positions) in the backbone between two residues required to define a “contact” is  $\langle n \rangle$ .

**--onlypdb**

Reads the structure from the pdbfile and ignores the alignment consensus structure (if provided).

**--draw\_nonWC**

Adds the non-canonical base pairs into the structure graphical output. For clarity, the default is to draw only the Watson-Crick base pairs. This option affects only the drawing of the structure. All base pairs (canonical or not) are used as part of the structure to perform the two-set statistical test.

Example of reading a structure from a PDB file for the FMN riboswitch:

```
> bin/R-scape --cntmaxD 4 --cntmind 3 --pdb tutorial/3f2q.pdb -s --onlypdb tutorial/RF00050.sto
```

This command line extracts contacts from the pdb file that are at a Euclidean distance  $\leq 4\text{\AA}$  in the PDB structure, and such that they are at least 3 residues apart in the backbone.

The output is

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 0.8.1 (Jul 2018)
# Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# -----
# Two-set statistical test (one test for annotated base pairs, another for all other pairs)
#
# Structure obtained from the pdbfile
# ij in alignment | ij in pdbsequence | basepair type
# 3 218 | 1 112 | WWc
# 4 216 | 2 110 | CONTACT
# 4 217 | 2 111 | WWc
# 4 218 | 2 112 | CONTACT
# 5 216 | 3 110 | WWc
# 5 217 | 3 111 | CONTACT
# 6 215 | 4 109 | WWc
# 6 216 | 4 110 | CONTACT
# .
# .
# .
# 192 202 | 87 96 | WWc
# 192 203 | 87 97 | CONTACT
# 193 198 | 88 92 | CONTACT
# 193 201 | 88 95 | WWc
# 193 202 | 88 96 | CONTACT
# 195 197 | 89 91 | CONTACT
# 195 198 | 89 92 | WHt
# 198 200 | 92 94 | CONTACT
# 198 201 | 92 95 | CONTACT
# 205 207 | 99 101 | CONTACT
# PDB: versions/rscape/rscape_v0.8/tutorial/3f2q.pdb
# contacts 169 (49 bpairs 35 wc bpairs)
# maxD 4.00
# mind 3
# distance MIN
# L 139
# alen 221
# pdblen 112
# ::[[[[[[[,,,,,<<<_____>>>,((((<<<<_____AA>>>>,<<<-----<_____>>>>,<<<<<_____>>>>>aa)AAAA----) ) a
```

```
# MSA RF00050_FMN.3f2q nseq 144 (144) alen 139 (221) avgid 69.18 (68.15) nbpairs 49 (0)
#
# Method Target_E-val [cov_min,conv_max] [FP | TP True Found | Sen PPV F]
# GTP 0.05 [-9.78,216.11] [1 | 14 49 15 | 28.57 93.33 43.75]
#
# left_pos right_pos score E-value
#-----
* 171 183 216.11095 1.6421e-10
* 170 184 211.69081 2.76699e-10
* 192 202 168.72417 4.95548e-08
* 8 213 149.71776 4.89982e-07
* 172 182 138.66664 1.84675e-06
* 169 185 137.23189 2.21548e-06
** 16 30 133.44999 3.53772e-06
* 5 216 131.02575 4.70876e-06
* 84 186 125.60806 9.0169e-06
* 17 29 112.04610 4.62895e-05
* 7 214 111.12654 5.13519e-05
* 6 215 96.43781 0.00029929
* 36 87 96.32752 0.00029929
* 94 163 78.81578 0.0024303
* 7 213 107.68588 0.0147937
```

All coordinates are relative to the input alignment. The annotation of all types of RNA base pairs (WWc, WWt, WHc,...) is produced by the program `rnaview` (Yang et al., 2003).

## Options for type of pairs tested

When performing the two-class statistical test (option `-s`) using a `pdfile` to read the structure, there are different options as to which types of base pairs are used to define the sample size for the base pairs test.

The options are:

### `--samplecontacts`

The basepair statistical test includes all the contacts identified in a PDB or/and as a RNA secondary structure included with a input alignment in Stockholm format. This is the default option for amino acid alignments if a PDB file is provided.

### `--samplebp`

For RNA alignments with only. The basepair statistical test includes base pairs of all 12 possible types. This is the default option for RNA/DNA alignments if a PDB file is provided.

### `--samplewc`

For RNA alignments only. The basepair statistical test includes only the canonical (Watson-Crick/Watson-Crick type) base pairs (A:U, G:C, G:U). This is the default option for RNA/DNA alignments if a consensus secondary structure is provided.

## Output options

### `--roc`

Produces a tabular output that provides statistics for each score value.

File `tutorial/updatedArisong.roc` looks like:

```
> more tutorial/updatedArisong.roc
```

```
# MSA nseq 95 alen 65 avgid 66.352419 nbpairs 20 (20)
# Method: GTP
#cov_score FP TP Found True Negatives Sen PPV F E-value
121.79543 0 2 2 20 2060 10.00 100.00 18.18 4.07104e-05
121.44018 0 2 2 20 2060 10.00 100.00 18.18 4.29443e-05
```

```

121.08494  0  2  2      20    2060      10.00 100.00  18.18  4.53006e-05
120.72970  0  2  2      20    2060      10.00 100.00  18.18  4.53006e-05
...

```

This file produces a tabular output for each alignment as a function of the covariation score, for plotting ROC curves. The values in the file are described by the comment line. Notice that the number of Trues (column 5) and Negatives (column 6) are fixed for a given secondary structure and do not change.

**--outmsa <f>**

The actual alignment analyzed can be saved in Stockholm format to file <f>.

**--outtree <f>**

The phylogenetic tree (created using the program FastTree) can be saved in Newick format to file <f>.

**--savenull**

Saves a histogram with the null distribution to file rnafile.msaname.null.

## Plotting options

**--nofigures**

None of the graphical outputs are produced using this option.

**--r2rall**

Forces R2R to draw all positions in the alignment. By default only those that are more than 50% occupied or are base paired are depicted.

## Other options

**--seed <n>**

Sets the seed of the random number generator to <n>. Use n = 0 for a random seed.

## 9 Some other topics

### How do I cite R-scape?

Rivas, E. *et al.*, “A statistical test for conserved RNA structure shows lack of evidence for structure in lncRNAs”, *Nature Methods* 14, 4548 (2017).

<https://www.nature.com/articles/nmeth.4066>

Rivas, E. and Eddy, S. E., “Response to Tavares *et al.*, Covariation analysis with improved parameters reveals conservation in lncRNA structures”, (2018).

<https://doi.org/10.1101/2020.02.18.955047>.

Rivas, E. *et al.*, “Estimating the power of sequence covariation analysis for detecting conserved RNA structure”, *Bioinformatics*, 36, 30723076, (2020).

<https://doi.org/10.1093/bioinformatics/btaa080>

Rivas, E., “RNA covariation at helix-level resolution for the identification of evolutionarily conserved RNA structure”, *PLOS Comput Biol*, 19(7): e1011262, (2023).

<https://doi.org/10.1371/journal.pcbi.1011262>.

### How do I cite CaCoFold?

Rivas, E., “RNA structure prediction using positive and negative evolutionary information”, *PLOS Comput Biol*, 16(10), e1008387, (2020).

<https://doi.org/10.1371/journal.pcbi.1008387>.

### How do I cite CaCoFold-R3D?

Karan, A and Rivas, E., “All-at-once RNA folding with 3D motif prediction framed by evolutionary information”, *bioRxiv*, Dec (2024).

<https://doi.org/10.1101/2024.12.17.628809>, 20 Dec 2024.

You should also cite what version of the software you used. We archive all old versions, so anyone should be able to obtain the version you used, when exact reproducibility of an analysis is an issue.

The version number is in the header of most output files. To see it quickly, do something like `R-scape -h` to get a help page, and the header will say:

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 2.5.7 (March 2025)
# Copyright (C) 2017-2025 Elena Rivas, Harvard University.
# Freely distributed under the GNU General Public License (GPLv3).
# -----
```

So (from the second line there) this is from R-scape v2.5.7.

### How do I report a bug?

Email us, at [elenarivas@fas.harvard.edu](mailto:elenarivas@fas.harvard.edu).

Before we can see what needs fixing, we almost always need to reproduce a bug on one of our machines. This means we want to have a small, reproducible test case that shows us the failure you’re seeing. So if you’re reporting a bug, please send us:

- A brief description of what went wrong.

- The command line(s) that reproduce the problem.
- Copies of any files we need to run those command lines.
- Information about what kind of hardware you're on, what operating system, and what compiler and version you used, with what configuration arguments.

## 10 Acknowledgments

We thank S.E. Roian Egnor for suggesting the name R-scape, and the Centro de Ciencias de Benasque Pedro Pascual in Spain, for their hospitality, over numerous and wonderful summers.

## References

- del Val, C., Romero-Zaliz, R., Torres-Quesada, O., Peregrina, A., Toro, N., and Jiménez-Zurdo, J. I. (2012). A survey of sRNA families in  $\alpha$ -proteobacteria. *RNA Biol*, 9:119–129.
- Dunn, S. D., Wahl, L. M., and Gloor, G. B. (2007). Mutual information without the influence of phylogeny or entropy dramatically improves residue contact predictions. *Bioinformatics*, 24:333–340.
- Fodor, A. A. and Aldrich, R. W. (2004). Influence of conservation on calculations of amino acid covariance in multiple sequence alignments. *Proteins: Structure, Function, and Bioinformatics*, 56(2):211–221.
- Gerstein, M., Sonnhammer, E. L. L., and Chothia, C. (1994). Volume changes in protein evolution. *J. Mol. Biol.*, 235:1067–1078.
- Gutell, R. R., Larsen, N., and Woese, C. R. (1994). Lessons from an evolving rRNA: 16S and 23S rRNA structures from a comparative perspective. *Microbiol. Rev.*, 58:10–26.
- Henikoff, S. and Henikoff, J. G. (1994). Position-based sequence weights. *J. Mol. Biol.*, 243:574–578.
- Hofacker, I. L., Fekete, M., and Stadler, P. F. (2002). Secondary structure prediction for aligned RNA sequences. *J. Mol. Biol.*, 319:1059–1066.
- Jung, S., Swart, E. C., Minx, P. J., Magrini, V., Mardis, E. R., Landweber, L. F., and Eddy, S. R. (2011). Exploiting *Oxytricha trifallax* nanochromosomes to screen for noncoding RNA genes. *Nucl. Acids Res.*, 39:7529–7547.
- Lindgreen, S., Gardner, P. P., and Krogh, A. (2006). Measuring covariation in RNA alignments: physical realism improves information measures. *Bioinformatics*, 22:2988–2995.
- Martin, L., Gloor, G., Dunn, S., and Wahl, L. (2005). Using information theory to search for co-evolving residues in proteins. *Bioinformatics*, 21:4116–4124.
- Nawrocki, E. P., Burge, S. W., Bateman, A., Daub, J., Eberhardt, R. Y., Eddy, S. R., Floden, E. W., Gardner, P. P., Jones, T. A., Tate, J., and Finn, R. D. (2015). Rfam 12.0: updates to the RNA families database. *Nucl. Acids Res.*, 43:D130–D137.
- Price, M. N., Dehal, P. S., and Arkin, A. P. (2010). FastTree 2 - approximately maximum-likelihood trees for large alignments. *PLOS ONE*, 5:e9490.
- Shannon, C. E. (1948). A note on the concept of entropy. *Bell System Tech. J.*, 27:379–423.
- Weinberg, Z. (2018). The Zasha Weinberg Database (ZWD). Available: <https://bitbucket.org/zashaw/zashaweinbergdata/>. Accessed 11 February 2019.
- Weinberg, Z. and Breaker, R. R. (2011). R2R – software to speed the depiction of aesthetic consensus RNA secondary structures. *BMC Bioinformatics*, 12:3.
- Weinberg, Z., Wang, J. X., Bogue, J., Yang, J., Corbino, K., Moy, R. H., and Breaker, R. R. (2010). Comparative genomics reveals 104 candidate structured RNAs from bacteria, archaea, and their metagenomes. *Genome Biol.*, 11(3):R31.
- Woolf, B. (1957). The log likelihood ratio test (the G-test). *Annals of Human Genetics*, 21:397–409.
- Yang, H., Jossinet, F., Leontis, N., Chen, L., Westbrook, J., Berman, H. M., and Westhof, E. (2003). Tools for the automatic identification and classification of RNA base pairs. *Nucl. Acids Res.*, 31.13:3450–3460.